

Antimicrobial activity of marine algae

M.Kausalya* and G.M. Narasimha Rao

Department of Microbiology*, Department of Botany, Andhra University, Visakhapatnam, Andhra Pradesh, India. *Corresponding Author: E-mail: kausalyasadamalla@gmail.com ; .9573921868

Abstract:

The present communication deals with the antimicrobial activity of the seaweeds such as *Sargassum polycystum* and *Sargassum tenerrimum* against both gram - positive, gram- negative and fungal pathogens. For experimental study different crude seaweeds extracts (Chloroform , Ethanol, Methanol and water) was determined by the well diffusion method. The inhibition zone was measured for all the Crude extracts revealed a wide range of antimicrobial activity against tested pathogens. Antimicrobial activity indicates that the presence of active constituents in the extractions of marine algae which can be exploited for the production of innovation drugs for the benefit of the humanity.

Keywords: Seaweeds, antimicrobial activity Well diffusion method.

Introduction

Among the marine flora and fauna marine algae are rich sources of diverse bioactive compounds with various biological activities. Recently, their importance as a source of novel bioactive substances is growing rapidly and researchers have revealed that marine algal originated compounds exhibit various biological activities (Wijesekara *et al.*, 2010). During the last years, many studies have been made on biological activities of the seaweed and identified as potentiala sources of natural antioxidants (Matanjun *et al.*, 2008). Several authors studied the antimicrobial activities of marine algae in different parts of our country (Battu *et al.*, 2011, Selvi *et al.*, 2011, Tuney *et al.*, 2006, Veeragurunathan and Geetha, 2009, Veeragurunathan *et al.*, 2008). In recent year seaweeds are wildly used in several applications such asantimicrobial (Chiheb *et al.*, 2011), antiviral (Bouhlal *et al.*, 2010, Bouhlal *et al.*, 2011), antifungal (De Felicio *et al.*, 2010), anti-allergic (Na *et al.*, 2005), anti-coagulant (Dayong *et al.*, 2008), anti-cancer (Kim *et al.*, 2011), anti-fouling (Bhadury and Wright, 2004) and antioxidant activities (Devi *et al.*, 2011). In the present study an attempt was made on antimicrobial properties of some marine occurring at mandapam coast.

Materials and Methods

Sample collection

Mandapam located on the southeast coast of Tamil Nadu (78°08 ' E and 9° 17' N) with luxuriant algal growth throughout the year . At Rameswaram the shore is sandy with boulders and platforms of compressed sandstones with rough end uneven surfaces situated at different level from high water to low water. Live and healthy marine algae were collected in the polythene bags with seawater and brought to the laboratory. Each species was washed with running water to remove epiphytes, animal castings, attached debris and sand particals, the final washings were done with distilled water and dried under shade.

Seaweeds extract preparation

This each Seaweed material mixed with different solvents with increasing polarity (Chloroform, Ethanol, Methanol and water) and placed into the Soxhlet apparatus. Each extraction was carried out in a Soxhlet apparatus for 24 hrs and after evaporation in vaccum the extracts were stored at -20°c until used (Krishnaveni *et ah*, 2012).

Bacterial and Fungal pathogens

For testing the antibacterial activity, the following Gram positive *Bacillus subtilis*(MTCC-441), *Staphylococcus aureus*(MTCC-96), *Micrococcus luteus*((MTCC-1538),*Staptococcus mutans*(MTCC-890),*Streptococcus anginosus*(MTCC-1929), *Lactobacillus acidophilus* (447) and Gram negative-*Escherichia coli, Enterobacter aerogenes*(MTCC-111), *Klebsiella pneumonia*(MTCC-432), *Pseudomonas aeuroginosa*(MTCC-424),*Erwinia caratovora* (MTCC-1428)and *Proteus vulgaris* (MTCC-1771) bacterial strains were selected. For antifungal activity, The following fungal strains, *Candida albicans*(MTCC-

227), Aspergillus niger(MTCC-1344), Saccharomyces cerevisiae(MTCC-463), Rhizoctonia solani(MTCC-984), Mucor racemosus(MTCC-6333) and Rhizopus stolonifer(MTCC-2198) were used for antifungal activity. They were obtained from the Institute of microbial technology Chandigarh. The work was carried out in Department of Microbiology, Andhra University.

Antimicrobial Activity by disc diffusion method:

In the present study, the antimicrobial activity of the seaweeds was studied by agar cup plate diffusion method (Kavangh, 1992). The Chloroform,Ethanol, Methanol and Water extracts of the collected test samples were tested in three dose levels of l00mg/ml, 300mg/ml, and 500mg/ml respectively. The nutrient agar medium prepared was inoculated with 18 hours old cultures of the above mentioned test organisms and were transferred into sterile 15cm diameter petridishes. The medium in the plates were allowed to set at room temperature for about 10 minutes and allowed to solidify in a refrigerator for about 30 minutes, 5 cups of 6mm diameter were made in each plate at equal distance. Stock solutions of the test residual extract were prepared in l00mg/ml, 300mg/ml, and 500mg/ml of each concentration were placed in the cups with sterile pipettes. In each plate one cup was used for control. Antibiotic Chloramphenical (l00mg/ml) was used as standard and respective solvents were used as control. The petridishes were prepared and incubated for 24 hrs at 37° C for bacteria. The above procedure is allowed for fungal assays but expects the media potato dextrose agar instead of nutrient agar and the antibiotic nystatin was used as standard. The plates were incubated at 25⁰c for 48hrs, after that the zone of inhibition was measured with zonal scale in mm and the experiment was carried out in duplicate.

Results

Antimicrobial activity of sargassum polycystum

Fig: 1.1 Shows the antimicrobial activity *Sargassum polycystum* at 100 mg/ml conc. of chloroform, ethanol, methanol and water extracts. In these four extracts ethanol and methanol showed moderate activity. Methanol extract showed a maximum activity against pathogens like *Proteus vulgaris* (18mm), *K. pneumonia* (18mm) and fungal strains methanol extract of *A.niger*(19mm), *R. stolonifer*(19mm), ethanol extract of *R. stolonifer*(19mm) showed promising activity.

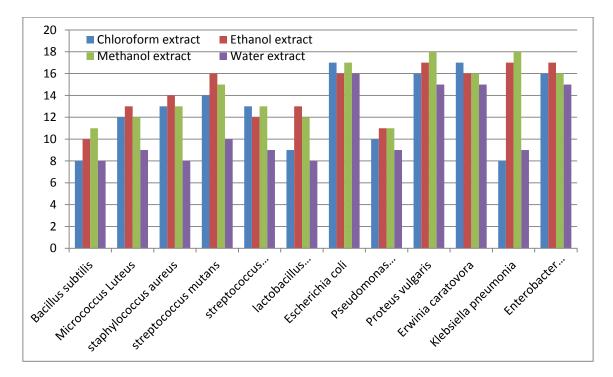


Fig: 1.1 Antibacterial activity of SARGASSUM POLYCYSTUM 100mg/ml

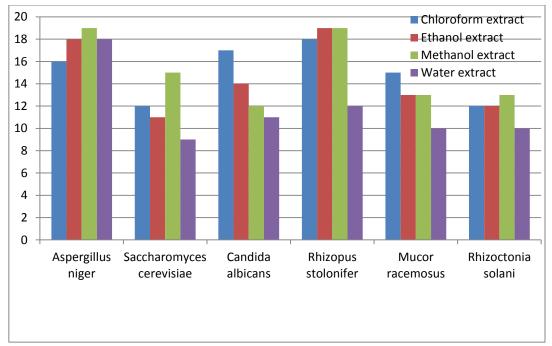


Fig: 1.1a Anti fungal activity of SARGASSUM POLYCYSTUM 100mg/ml

Fig:1.2 Shows the antimicrobial activity Sargassum polycystum in 300mg/ml conc. of chloroform, ethanol, methanol and water extracts. In these four extracts methanol and ethanol extract showed moderate activity. Methanol extract of E.coli(19mm), P.vulgaris (19mm), K. pneumonia (19mm), ethanol extract P.vulgaris (19mm), E.aerogenes(19mm) showed promising activity and fungal strain of methanol extract R. stolonifer(21mm) showed high activity and Water extract of S.cerevisiae showed low activity.

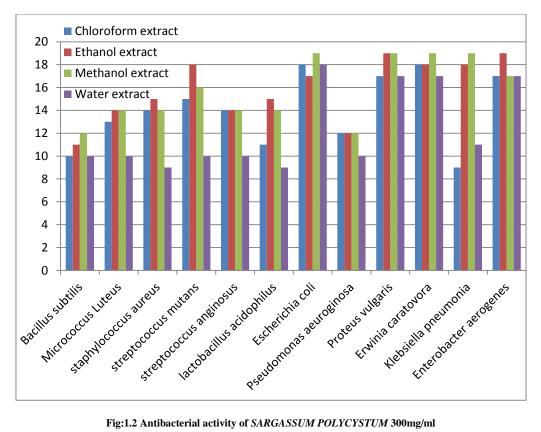


Fig:1.2 Antibacterial activity of SARGASSUM POLYCYSTUM 300mg/ml

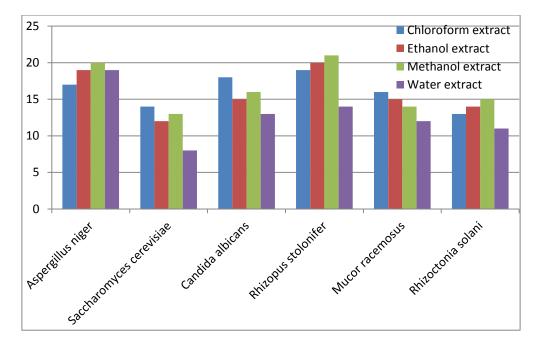


Fig:1.2aAnti fungal activity of SARGASSUM POLYCYSTUM 300mg/ml

Fig: 1.3. Shows of antimicrobial activity *Sargassum polycystum* at 500mg/ml conc. of chloroform,ethanol, methanol and water extracts. In these four methanol extract showed high activity. The extract obtained using chloroform showed maximum activity against pathogens like *E.coli*(19mm), *E. caratovora*(19mm), *E.aerogenes*(19mm) and showed low activity against *K. pneumonia* (11mm). Fungal stains *A.niger*(20mm), *R. stolonifer*(21mm) and *Candida albicans*(20mm) showed high activity. Ethanol extract observed the highest activity against pathogens like *S. aureus* (20mm), *K. pneumonia* (21mm) *P.vulgaris* (20mm), *E. caratovora*(20mm), *E.aerogenes*(20mm), *R. stolonifer*(21mm), *C. Candida albicans*(20mm) showed high activity. The extracts obtained using methanol showed highest activity against pathogens like *S. aureus* (20mm), *K. pneumonia* (21mm), *P.vulgaris* (20mm), *B. caratovora*(20mm), *K. pneumonia* (21mm), *P.vulgaris* (20mm), *B. caratovora*(21mm), *K. pneumonia* (21mm), *A.niger*(22mm), *R. stolonifer*(22mm). The extracts obtained using water showed highest activity against pathogens like *E.coli*(19mm), *P.vulgaris* (19mm), *B. caratovora*(21mm), *K. pneumonia* (21mm), *A.niger*(22mm), *R. stolonifer*(22mm). The extracts obtained using water showed highest activity against pathogens like *E.coli*(19mm), fungal stains *A.niger*(20mm) showed highest activity and extract of *S.cerevisiae* showed mild activity.

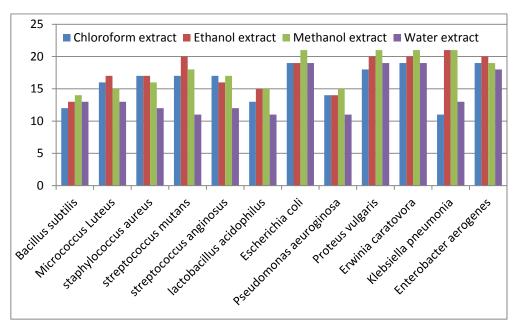


Fig: 1.3 Antibacterial activity of SARGASSUM POLYCYSTUM 500mg/ml

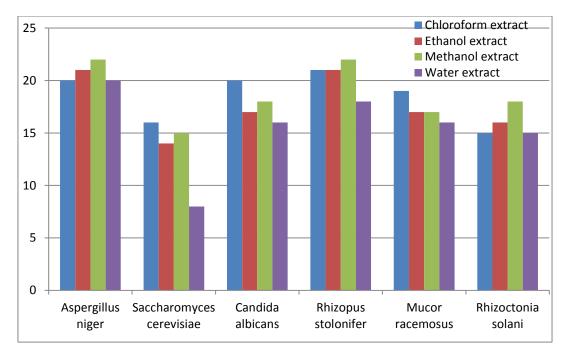


Fig: 1.3a Anti fungal activity of SARGASSUM POLYCYSTUM 500mg/ml

Antimicrobial activity of Sargassum tenerrimum

Fig: 2.1 .Shows the antimicrobial activity of *Sargassum tenerrimum* at 100 mg/ml conc. of chloroform ethanol, methanol and water extracts. In these four extracts ethanol and chloroform showed moderate activity. Ethanol extract shows promising acyivity against pathogen like of *S. aureus*(15mm) and fungal strains A.niger(10mm), *R. stolonifer*(10mm)and *M.racemosus*(10mm) showed mild activity.

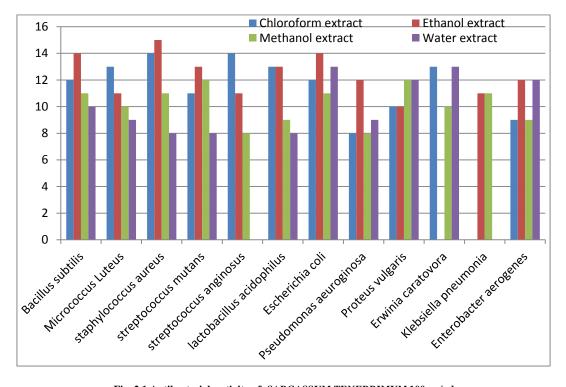


Fig: 2.1 Antibacterial activity of SARGASSUM TENERRIMUM 100mg/ml

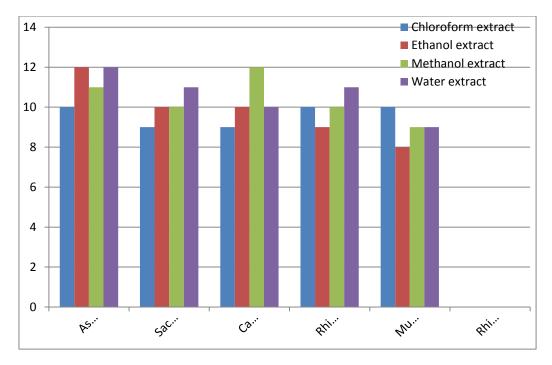


Fig: 2.1a Antifungal Activity of SARGASSUM TENERRIMUM 100mg/ml

Fig: 2.2 Shows the antimicrobial activity of *Sargassum tenerrimum* at 300mg/ml conc. of chloroform,ethanol,methanol and water extracts. Among these four extracts ethanol extract showed highest activity against pathogen like *S. aureus* (19mm) and fungal strain of water extracts of A.niger(14mm), methanol extract of *Candida albicans*(13mm)and ethanol extract of A.niger(13mm) showed moderate activity

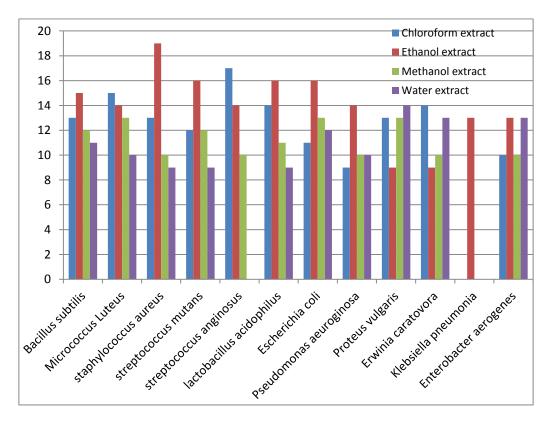


Fig: 2.2 Antibacterial activity of SARGASSUM TENERRIMUM 300mg/ml

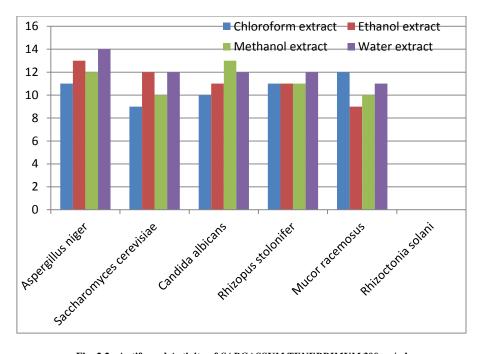


Fig: 2.2a Antifungal Activity of SARGASSUM TENERRIMUM 300mg/ml

Fig:2.3. Shows the antimicrobial activity of *Sargassum tenerrimum* at 500mg/ml conc. of chloroform, ethanol, methanol and water extracts. In these four extracts ethanol and chloroform extracts showed moderate activity. Chlorofrom etract showed promising activity against pathogens like *S. aureus* (17mm), *S.anginosus*(18mm), and *M.racemosus*(13mm) showed mild activity. Ethanol extract proved the highest activity against pathogens like *S. aureus* (17mm), *S. carevisiae* (13mm) and fungal strain *A.niger*(14mm), *S. cerevisiae* (13mm) showed mild activity. Methanol extract observed the promising activity against pathogens like *E. coli*(15mm), *P. vulgaris* (15mm), *Candida albicans*(14mm) and *R. Solani*(10mm) showed low activity. Water extract observed the promising a activity against pathogens like *E. coli*(14mm), *P. vulgaris* (14mm) and *A.niger*(15mm), *R. stolonifer*(14mm), showed moderate activity.

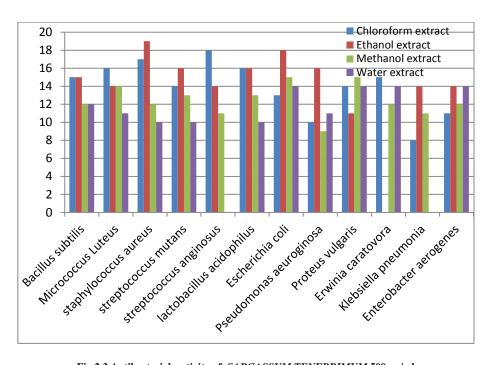


Fig:2.3 Antibacterial activity of SARGASSUM TENERRIMUM 500mg/ml

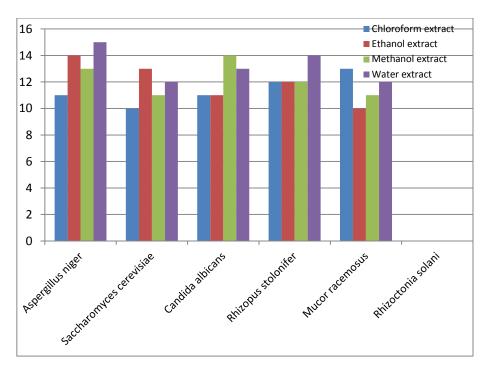


Fig: 2.3a Antifungal Activity of SARGASSUM TENERRIMUM 500mg/ml

Discussion

The present study was to evaluate the antimicrobial activity of the different macroalgae for its bioactive potentials. Rodriguez et al., (2010), Bhacuni and Rawat, (2005), Priyadharshini et al,(2011) have reported that seaweeds are an excellent source of components such as polysaccharides, tannins, flavonoids, phenolic acids, bromophenols, and carotenoids has exhibits different biological activities. Depending upon their solubility and polarity, different solvents shows the different antimicrobial activity. So chemical compounds should be extracted from different seaweeds in order to optimize their antibacterial activity by selecting the best solvent system (Hediat et al.,). The results from the present study revealed that the strongest antibacterial activity exhibited by the methanol extract by Sargassum polycystum .Methanol have higher antibacterial activity than that of extracts obtained with other organic solvents (Febles et al., 1995; Sidharata et al., 1997; Kumar et al., 2008; Seenivasan et al.,2010;Lavany and veerappan,2011).Devi et al.,2008;Meenakshi et al.,2009;Cox et al.,2010;Srivastava et al.,2010, reported that the methanol extract of seaweeds contains phenolics, alkaloids and amino acids which may responsible for the antimicrobial activity. Manilal et al., and Rangaiah et al., exlained that the methanol extraction yielded higher antimicrobial activity than n-hexane and ethyl acetate . Methanol extract of Sargassum polycystum showed more activity against E.coli, P.vulgaris, E. caratovora, K. pneumonia and fungal strain of methanol extract of A.niger and R. stolonifer. Chloroform extract showed moderate activity with Sargassum tenerrimum. Ethanol extract of Sargassum tenerrimum showed highest activity against S.aureus and water extract of A.niger showed moderate activity. Water extract shows less activity Hodgson(1944). The result shows the activity of four solvent extracts in the order methanol > ethanol > chloroform > aqueous extracts agreeing with Vizaya parthasarathy et.al.,(2004). The present study reveals that solvents are always better for extraction when compared with water.

Conclusion

The results of this present study on *sargassum polycystum* and *sargassum ternerrimum* using four different solvent extract against eighteen different pathogens showed significant antimicrobial activity. However, more research has to be done on isolation ,purification and identification of the active ingredients in order to understand their bio prospects.

References

Battu GR, Ethadi S, Prayaga Murthy P, Praneeth VS, Rao M. (2011). In- vitro antibacterial activity and preliminary phytochemical screening of three algae from Visakhapatnam Coast, Andhra Pradesh, India. Int. J. Pharm. Sci : 3 (4): 399-401.

Bhacuni, D.S. - Rawat, D.S. (2005). *Bioactive Marine Natural Products*. Spinger/Anamaya Publishers. 400p. ISBN: 978-1402034725.

Bhadury, P. Wright, C.P. (2004). Exploitation of marine algae: biogenic compounds for potential antifouling application. In Planta, vol. 219 : p.561-578.

Bouhlal, R.- Riadi, H. Bourgougnon N. (2010). Antiviral Activity of the extracts of Rhodophyceae from Morocco. In African Jornal of Biotecnology, vol. 9: p.7968-7975.

Bouhlal, R.- Haslin, C- Chermann, J.C.- Colliec- Jouault, S.- Sinquin, C- Simon, G. - Cerantola, S.- Riadi, H-Bourgougnon, N. (2011). Antiviral activities of sulfated Polysaccharides isolated from *Sphaerococcus coronopifolius* (*Rhodophyta, Gigartinales*) and Boergeseniella thuyoides (*Rhodophyta, Ceramiales*). In Marine Drugs, vol. 9: p. 1187-1209.

Chiheb, I. Riadi, H. Martine -Lopez, J. Dominguez-Seglar, J.F. Gomez-Vidal, J.A. Bouziane, H. Kadiri, M. (2009). Screeening of antibacterial activity in marine green and brown macroalgae from the coast of Morocco. *In* African Journal Of Biotechnology, vol. 8, p. 1258-1562.

Cox, S., Abu-Ghannam, N. and Gupta, S. 2010. An assessment of the antioxidant and antimicrobial activity of six species of edible Irish seaweeds. International Food Research Journal, 17: 205-220.

Dayong, S.-Jing, L- Shuju, G.-Lijun, H. (2008). Antithrombotic effect of bromophenol, the alga- derived thrombin inhibitor. In Journal of Biotechnology, vol.136: p.763-769.

De Felicio, R- De Albuquerque, S.-Young, M.C.M.-Yokoya, N.S-Debonsi, H.M. (2010). Trypanocidal, leishmanicidal and antifungal potential from marine red alga Bostrychia tenella, J.Agardh (Rhodomelaceae, Ceramiales). In Journal Of Pharmaceutical And Biomedical Analysis, vol. 52: p. 763-769.

Devi, G.K.Manivannan, K. Thirumaran, G. Rajathi, F.A.A. Anantharaman, P. (2011). In vitro antioxidant activities of selected Seaweeds from southeast coast of India. In Asian Pacific Journal Of Tropical Medicine, vol. 4: p. 205-211.

Devi, K.P., Suganthy, N., Kesika, P. and Pandian, S.K. 2008. Bioprotective properties of seaweeds: in vitro evaluation of antioxidant activity and antimicrobial activity against food borne bacteria in relation to polyphenolic content. BMC Complementary and Alternative Medicine, 8: 38-48.

Febles, C.I., Arias, A., Gil-Rodriguez, M.C., Hardisson, A. and Sierra Lopez, 90 K.J. Elnabris, A.A. Elmanama and W.N. Chihadeh A. 1995. In vitro study of antimicrobial activity in algae (Chlorophyta, Phaeophyta and Rhodophyta) collected from the coast of Tenerife (in Spanish). Anuario del Estudios Canarios, 34: 181-192.

Hediat MH. Salama, Najat Marraiki. (2010). Antimicrobial activity and phytochemical analyses of Polygonum aviculare L. (*Polygonaceae*), naturally growing n Egypt. Saudi Journal Of Biological Sciences: 17. 57-63.

Hodgson, L.M. 1984. Antimicrobial and antineoplastic activity in some south florida seaweeds. Botantica Marine .28:387-390.

Kavangh F. (1992). Analytical Microbiology-II, Academic press, New York: 241-243.

Kim SK, Wijesekara I. Development and biological activities of marine derived bioactive peptides: A review. Journal of Fuctional Foods 2010; 1 -9.

Kim, S.K.- Karadeniz, F. (2011). Anti-HIV Activity of extracts and compounds from marine algae. In Advanced Food and Nutrition Research, vol. 64: p.213-224.

Kim SK, THOMAS, N.V. - LI, X. (2011). Anticancer compounds from marine macroalgae and their application as medicinal foods. Advanced Food and Nutrition Research, vol. 64,2011, p. 213-224.

Kolanjinathan K, Ganesh P, Govindarajan M. (2009). Antibacterial activity of Ethanol extracts of seaweeds against fish bacterial pathogens. Eur.Rev. Med.Pharmacol.Sci :13:173-177.

Krishnaveni Eahamban, Johnson Marimuthu Antonisamy. (2012). Preliminary Phytochemical, UV-VIS, HPLC and Antibacterial studies on *Gracilaria cirticata* J. Ag. Asian Pacific Journal of Tropical Biomedicin: 2012. S568-S574.

Kumar, G.S., Sarada, D.V.L. and Rengasamy, R. 2008. Seaweed extracts control the leaf spot disease of the medicinal plant Gymnema sylvestre Indian Journal of Science and Technology, 1(3): 1-5.

Lavanya, R. and Veerappan, N. 2011Antibacterial Potential of six seaweeds collected from Gulf of Mannar of Southeast Coast of India. Advances in Biological Research, 5(1): 38-44.

Manilal A, Sujith S, Selvin J, Shakir C, Kiran GS. (2009). Antibacterial activity of Falkenbergia hillebrandii (Born) from the India coast against human pathogens. FYTON 78:161-166.

Manjula E and Narasimha rao(2014) Invitro study of antimicrobial activity of marine algae *caulerpa taxifolia* and *caulerpa Racemosa* (C.Agardh), IJABPT vol-5, issue-2(pg:57-62).

Matanjun P, Mohamed S, Mustapha NM, Mohammad K, Ming CH. (2008). Antioxidant activities and phenolics content of eight species of seaweeds from north Borneo. J.Appl. Phycol: 20(4): 367-373.

Meenakshi, S., Manicka Gnanambigai, D., Tamil mozhi, S., Arumugam, M. and Balasubramanian, T. 2009. Total flavanoid and in vitro antioxidant Antibacteria from seaweeds of Gaza 91 activity of two seaweeds of Rameshwaram coast. Global Journal of Pharmacology, 3 (2): 59-62.

NA, H.J.-Moon, P.D.-Lee, H.J.-Kim, H.R-Chae, H.J.-Shin, T. -Seo, Y.-Hong, S.H-Kim, H.M. (2005). Regulatory effect of atopic allergic reaction by Corpopelt is affinis. In Journal of Ethnopharmocology, vol.101: p.43-48.

Priyadharshini, S.-Bragadeeswaran, S.- Prabhu, K - Ran, S.S. (2011). Antimicrobial activity and hemolytic activity of seaweed extracts *Ulva fasciata* (Delile 1813) from Mandapam, Southeast coast of India. In Asian Pacific Journal of Tropical Biomedicine, vol.1, 2011, p.S38-S39.

Rajasulochana P, Dhamotharan R, Krishnamoorthy P, Murugesan S. (2011). Antibacterial activity of the extracts of marine red and brown algae. J.Am.Sci : 3(4):399-401.

Rangaiah GS, Lakshmi PA, Sruthi keerithia K. (2010). The antimicrobial activity of the crude extracts of *Chlorophyceacan* Seaweeds *Ulva, Caulerpa* and *Spongontorpha* spp. against clinical and phyto pathogens. Drug Invent. To day :2:311-314.

Rodriguez- Bernaldo de Quiros A, Large- Yusty MA, Lopez- Hernadez J. (2010). Determination of phenolic compounds in macroalgae for human consumption. Food Chem : 121. 634-638.

Selvi M.R. Selvaraj and Anandhi Chidambaram. (2001). Screening for antibacterial activity of macroalgae. Seaweed Res.Utilin, 23: (1&2). 59-63.

Seenivasan, R., Indu, H., Archana, G. and Geetha, S. 2010. The Antibacterial Activity of Some Marine Algae from South East Coast of India. American- Eurasian Journal of Agricultural & Environmental Science, 9(5): 480-489.

Sidharta, A.K., Mody, K.H., Ramavat, B.K., Chauhan, V.D., Garg, H.S., Goel, A.K., Doss, M.J., Srivastava, M.N., Patnaik, G.K. and Kamboj, V.P. 1997.Bioactivity of marine organisms: Part VIII-Screening of some marine flora of western coast of India. Indian Journal of Experimental Biology, 35(6): 638-643.

Srivastava, N., Saurav, K., Mohanasrinivasan, V., Kannabiran, K. and Singh, M. 2010. Antibacterial potential ofmacroalgae collected from the Madappam coast, India. British J. Pharm. & Toxicol., 1(2): 72-76.92 K.

Tuney, I., B.H. Cadirci, D.Unal and A. Sukatar. (2006). Antimicrobial activities of the extracts of marine algae from the coast of Ural (Izmir Turkey) Brittany France. TurkJ.Biol :30: 171 -175.

Veeragurunathan. V and T. Geetha. (2009). Screening for antimicrobial activity of marine algae from Gulf of Mannar. Tamil Nadu. Seaweed Res.Utili: 31(1&2).151-155.

Veeragurunathan. V., R. Vellaiyan, S.Siva Subramanian, G.Sadannantham and R. Bahkyaraj. (2008). Studies on the antibacterial activity of selected seaweeds from Gulf of Mannar. Seaweed Res.Utilin: 30(1 & 2). 147-152.

Vijaya parthasarathy ,M.D,Prema, M and Krishnamurthy, A.(2004)Preliminary report on the antibacterial activity of extracts from spirogyra sp. Zygnematacea-chloophyceae.Indian Hydrobiology.7:229233.

Wijesekara I, Kim SK. (2010). Angiotensin-I-converting enzyme (ACE) inhibitors from marine resources: Prospects in the pharmaceutical industry. Marine Drugs :8:1080-1093.