



## Table of Contents

### Journal Articles

**Pathogenicity of *Leptospira icterohaemorrhagiae* serovar *Lai* strain Langkawi in Guinea Pigs (*Cavia porcellus*)**  
*H. Tyagita, A.R. Bahaman, S. Jasni, T.A.T. Ibrahim and N.H. Fuzina*.....1

**Antibiotics susceptibility of *Staphylococcus aureus* and *Escherichia coli* isolated from dairy goats in selected farms in Selangor, Malaysia**  
*R. Mansor, N.S. Diauddin, S.S. Syed-Hussain and S.F. Khalid*.....12

**Evaluation of leptospirosis knowledge, attitude and practice among dog handlers**  
*S.H. Goh, K.H. Khor, S.F. Lau, R. Ismail, S. Khairani-Bejo and R. Radzi*.....17

**Concentration of serum amyloid A in a clinically normal endurance horses in Malaysia**  
*S.K. Rajendren, N.H. Khairuddin, and S. Sumita* .....28

### Case Reports

**Lesson learnt: First reported case of complicated cutaneous pythiosis in a dog in Malaysia**  
*S. Omar, S.F. Lau, N.A.I. Mohd Zainudin, N. Azman and J.X. Jessie-Bay*.....34

**Dynamic cervical lung lobe herniation in a Shih Tzu dog**  
*T.S.Y. Joanne, M.L. Lim, O. Asnawi, M.A.H. Shamsul and K.H. Khor*.....38

### Short Communications

**Tenderness and cooking loss in meat of male Dorper Sheep weaned at different ages**  
*Z. Norhayati, M. Wan Zahari, S. Shanmugavelu and A. Dzulfazly*.....43

**Acknowledgements**.....45

**Guidelines for authors**.....46

**List of abbreviations and symbols**.....49

## EDITORIAL BOARD 2018-2020

### Editor-in-Chief:

Dr. Khor Kuan Hua

### Editors:

Prof. Dato' Dr. Abdul Rani Bahaman  
Prof. Dr. Saleha Abdul Aziz  
Prof. Dr. Rasedee Abdullah  
Prof. Dr. Goh Yong Meng  
Prof. Dr. Latiffah Hassan  
Assoc. Prof. Dr. Sharifah Syed Mohd Hassan  
Assoc. Prof. Dr. Chen Hui Cheng  
Assoc. Prof. Dr. Faez Firdaus Jesse Abdullah  
Assoc. Prof. Dr. Lau Seng Fong  
Dr. Ong Bee Lee  
Dr. Rozanaliza Radzi  
Dr. Sharifah Salmah Syed Hussain  
Dr. Sandie Choong Siew Shean  
Lt. M. Dr. John Shia Kwong Siew  
Dr. Hasliza Abu Hassim  
Dr. Azlan Che Mat  
Dr. Nik Mohd Faiz Nik Mohd Azmi

### Technical Support:

Dr. Goh Soon Heng

## EDITORIAL ADVISORY BOARD 2018-2020

Prof. Dato' Dr. Mohd. Hair Bejo  
University Putra Malaysia

Dato' Dr. Quaza Nizamuddin Bin Hassan Nizam  
Department of Veterinary Services  
Malaysia, Putrajaya

Prof. Datin Paduka Dato' Dr. Aini Ideris  
Universiti Putra Malaysia

Prof. Dr. Husni Omar Mohamed  
Cornell University, USA

Dr. Chandrawathani Panchadcharam  
Department of Veterinary Services  
Malaysia, Putrajaya

Prof. Dr. Jasni bin Sabri  
Universiti Malaysia Kelantan

Prof Dr. Paul Mills  
The University of Queensland,  
Australia

Prof Dr. Heng Hock Gan  
Purdue University, USA

Prof Dr. Huang Hui-Pi  
National Taiwan University, Taiwan

Prof. Dr. Mohd Azam Khan bin Goriman Khan, Universiti Malaysia Kelantan

## CONCENTRATION OF SERUM AMYLOID A IN CLINICALLY NORMAL ENDURANCE HORSES IN MALAYSIA

S.K. Rajendren<sup>1</sup>, N.H. Khairuddin<sup>2\*</sup>, and S. Sumita<sup>3</sup>

<sup>1</sup> Faculty of Veterinary Medicine, Universiti Malaysia Kelantan, Kota Bharu, Kelantan, Malaysia

<sup>2</sup> Faculty of Veterinary Medicine, Universiti Putra Malaysia, UPM Serdang, Selangor Darul Ehsan, Malaysia

<sup>3</sup> Faculty of Agriculture, Universiti Putra Malaysia, UPM Serdang, Selangor Darul Ehsan, Malaysia

### SUMMARY

Endurance horses continuously undergoing training. This will cause inflammation which leads to acute phase reaction with the production of acute phase protein, especially serum amyloid A (SAA). The purpose of this study was to establish concentration of SAA in normal endurance horses in the blood serum using two-site enzyme linked immunoassay (ELISA) technique. Horse sera were aliquoted from blood taken from jugular venipuncture. The highest concentration of SAA was observed in horses rested between 12 months and 24 months. The lowest concentration of SAA was noticed in horses rested more than 24 months. All the horses between 6 and 11 years old have high SAA concentration. When resting intervals were compared against gender of the horses, it was noted that all mares have high SAA concentration compared to gelding and stallion. Whereas SAA concentration in Thoroughbred horses were high compared to Arabian horses in all rest intervals. The SAA concentration in horses rested more than 24 months was low most probably because the horses recovered well from the inflammatory process happened during the endurance race.

Keywords: Serum Amyloid A, endurance horse, training, age, rest interval, Malaysia

### INTRODUCTION

Although long distance horse racing began in 1892, the first modern endurance ride was held in 1955. That race was a one-day event in which the horses compete in a 100-mile (160km) race from Nevada to California, United States of America (Nagy *et al.*, 2012). In 1966, Australia held the 160km Tom Quilty Gold Cup and this event is considered the second oldest modern horse endurance race. However, there are records that show Spain and Portugal have held conducting endurance horse rides since the 1950s. The European Long Distance Rides Conference (ELDRIC) was formed later in 1979 and under auspices of ELDRIC organised and regulated races were conducted in the North American, Australian and South Africa. The sports developed further when the endurance races became a Fédération Equestre Internationale (FEI) discipline in 1982 and subsequently most international rides are sanctioned and governed by FEI regulations (Nagy *et al.*, 2012). The endurance races are categorised according to distance, which are 40, 80, 120 and 160km races. Since these endurance rides can be potentially hazardous to the health of the competing horses, the races are punctuated with compulsory stops, during which the horses are inspected by veterinarians at the veterinary gate to ensure that the horses are fit to continue to the next phase. Due to the nature of the sport, competing horses are prone to develop career-limiting or even worse, fatal metabolic and orthopaedic disorders. Thus in these races, horses are eliminated from the ride if the metabolic status or orthopaedic conditions show that they are not fit to continue.

The acute phase proteins (APPs) are known indicators of inflammation (Pepys *et al.*, 1983; Kent 1992;

Gabay *et al.*, 1999; Cywińska *et al.*, 2012). The APPs are either positive indicators that increase or negative indicators that decrease in the presence of inflammation (Pepys *et al.*, 1983; Aldred and Schreiber, 1993; Gabay *et al.*, 1999). Positive APPs further divided into major APPs, which can increase to 10 to 1000 folds and in the presence of inflammatory triggers and minor APPs, which increase minimally in the inflammations and infections (Pepys *et al.*, 1983; Kent 1992; Kushner & Mackiewicz, 1993). In horses, the only major positive APPs is serum amyloid A (SAA) (Mozes *et al.*, 1989).

SAA is synthesised in the liver and its production ceases immediately upon recovery from inflammatory diseases or conditions (Uhlir and Whitehead, 1999). The basal serum concentration is low and the reference range of SAA is narrow (Kent, 1992; Koj *et al.*, 1996).

Equine athletes, especially endurance horses, due to the nature of their activities are prone to the development of inflammatory conditions, either through injuries or diseases. Currently, there is no reference values on SAA in endurance horses that could be used to determine the presence of inflammatory diseases and conditions in these horses. Thus, the aim of this research is to determine the level of SAA in competition endurance horses during their resting interval between races to develop a reference range and baseline for SAA in clinically healthy resting endurance horses.

### MATERIALS AND METHODS

#### Horses

The study was conducted on 40 endurance horses from Selangor and Terengganu, Malaysia. All the horses were either Arabians or Thoroughbreds comprising of 20 geldings, 16 mares, and 4 stallions, with ages ranging from 6 to 22 years. All these horses had participated in endurance competitions within the period of 6 to 43

\*Corresponding author: Assoc. Prof. Dr. Nurul Hayah Khairuddin (N.H. Khairuddin); Email: [nurulhayah@upm.edu.my](mailto:nurulhayah@upm.edu.my)

months prior to this study. Horses included in the study were undergoing training. The horses were clinically normal based on history and physical examination. The age, breed, and gender of the horses were recorded.

*Blood Samples*

Blood samples were taken from the jugular vein of horses while in stalls. The ambient temperature and environment at time of sampling were recorded. The sampling area of the jugular vein was occluded and swabbed with alcohol to sterilise. Blood samples were collected in two 3 mL plain tubes and allowed to clot for 20 minutes before centrifuging at  $3400 \times g$  for 10 minutes. Serum was separated and transferred to Eppendorf™ 1 mL tubes and stored at  $-20^{\circ} C$ . The SAA levels were determined by the double sandwich ELISA.

*Serum Amyloid A*

Determination of the SAA was conducted using the ELISA kit (Cusabio, Immunology Consultants Laboratory, USA). The serum samples were first diluted to 1/200 with by mixing 2  $\mu L$  of serum sample with 398  $\mu L$  of 1X diluent and the mixture mixed thoroughly. All reagents for the assay were brought into room temperature before use. 100  $\mu L$  of blank and each standard solution of 2.25, 4.5, 9, 18, 36, and 72 ng/mL SAA were pipetted in duplicate into the respective well of the 96-well ELISA microtiter plate. One hundred microlitres of diluted samples were pipetted into the designated wells. The plate was covered placed on a level surface and incubated while shaking, on an orbital shaker at room temperature for 60 minutes. The contents of the wells were then aspirated and the plate washed with wash solution provided in the kit. One hundred microlitres of diluted antibody solution were pipetted into each well and the plate were covered, placed on a level surface, and incubated while shaking on an orbital shaker at room temperature for 20 minutes in a dark room. The contents of the wells were aspirated and the plate washed wash solution and the wells blotted to remove moisture. One hundred microlitres of diluted HRP-streptavidin was pipetted into each well and the plate similar incubated while shaking on an orbital shaker at room temperature for 20 minutes in a dark room. The contents of the wells were aspirated and the washed wash solution, and the plate blotted. One hundred microlitres of TMB substrate solution was pipetted into each well and the plate incubated while shaking on an orbital shaker at room temperature for 10 minutes in a dark room. The reaction was terminated by adding 100  $\mu L$  of stop solution to each well. The absorbance (450nm) for the contents of the wells were spectrophotometrically determined according to manufacturer’s specifications.

*Statistical Analysis*

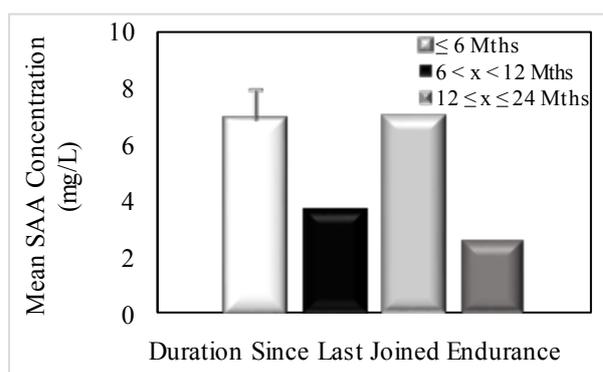
Statistical analysis, means and standard errors of mean were computed using SPSS® 20 for Windows® Microsoft. The results are expressed as the mean  $\pm$  standard error of the mean (SEM). The SAA

concentrations were compared among age group, gender, and breed of horses. Significance differences among means were determined at  $\alpha = 0.05$ .

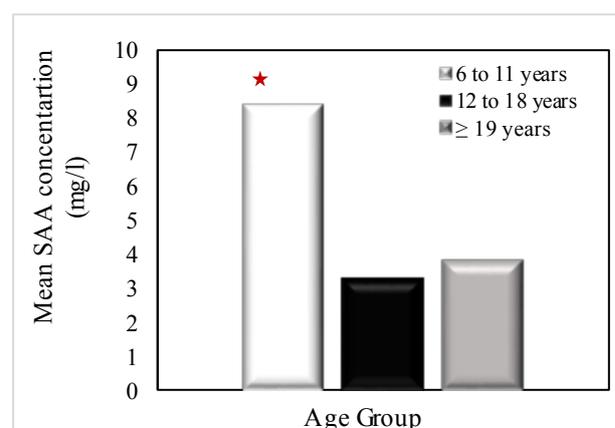
**RESULT**

There were a total of 40 horses included in the study and the data from all these horses were analysed and presented here. Among the 40 endurance horses in this study 16 (40%) had participated in endurance race with the period  $\leq 6$  months, 14 (35%) 6 to 12 months, 4 (10%) 12 to 24 months, and 6 (15%)  $> 24$  months prior to the study. The subjects comprised of 14 horses (35%) aged 6 to 11 years, 20 (50%) 12 to 18 years, and 6 (15%)  $> 19$  years old. 36 were Arabian horses (90%) and 4 Thoroughbreds (10%) of which 20 (50%) were geldings, 16 (40%) mares and 4 stallions (10%).

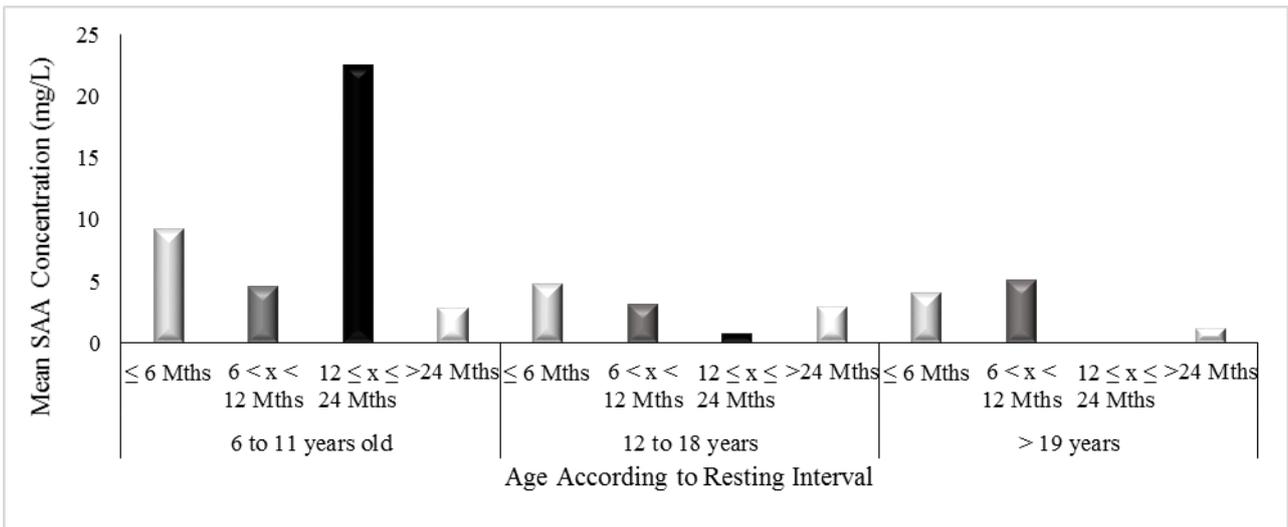
Based on previous participation in endurance races, the mean serum amyloid A (SAA) concentrations were highest in horses that participated 12 to 24 months ( $7.22 \pm 5.22$  mg/L) followed in order by horses that participated  $\leq 6$  months ( $6.87 \pm 1.74$  mg/L), 6 to 12 months ( $3.71 \pm 0.44$  mg/L) and  $> 24$  months ( $2.62 \pm 0.30$  mg/L) before the study (Figure 1).



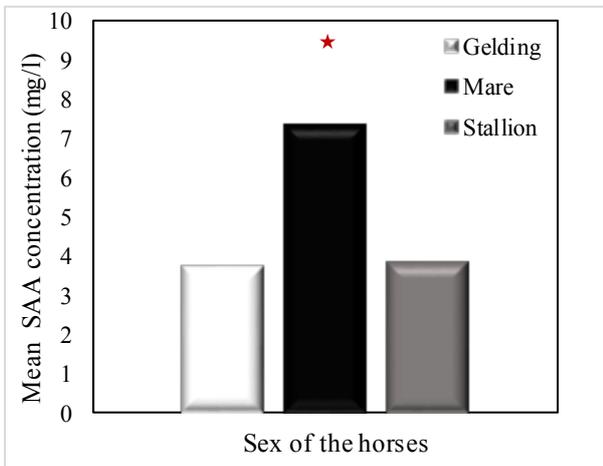
**Figure 1. Comparative mean SAA concentration (mg/L) of clinically normal endurance horses (n = 40) at four periods from three stables**



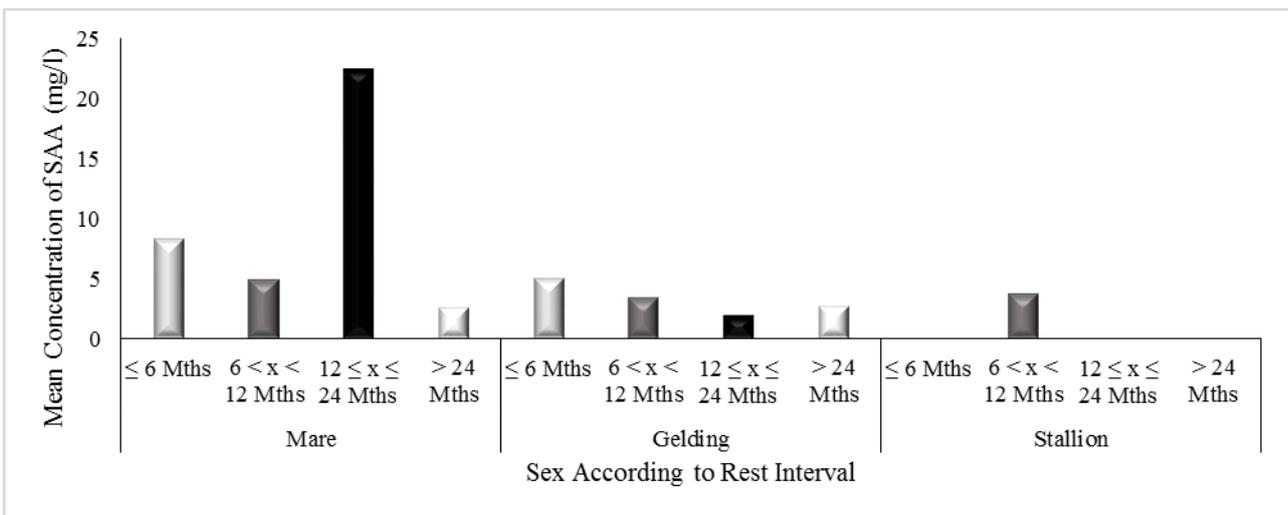
**Figure 2. Comparative mean SAA concentration (mg/L) of clinically normal endurance horses (n = 40) at three age group from three stables. The ‘★’ indicate significant mean difference**



**Figure 3. Comparative mean SAA concentration (mg/L) of clinically normal endurance horses (n = 40) at three age group according to rest interval from three stables**



**Figure 4. Comparative mean SAA concentration (mg/l) between clinically normal endurance geldings (n = 20), mares (n = 16) & stallion (n = 4)**



**Figure 5. Comparative mean SAA concentration (mg/L) between clinically normal endurance geldings (n = 20), mares (n = 16) & stallions (n = 4) according to rest interval**

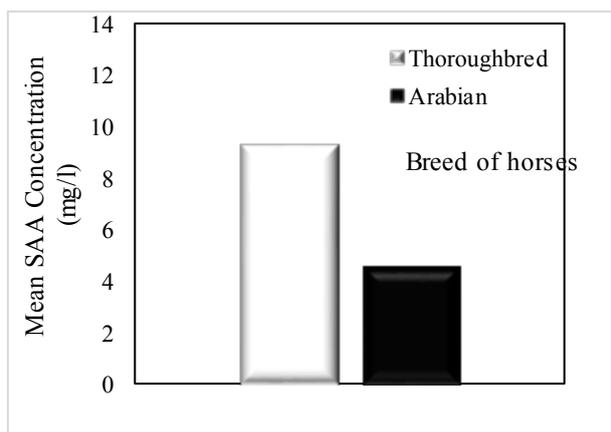


Figure 6. Comparative mean SAA concentration (mg/L) between Thoroughbred (n = 4) and Arabian (n = 36) endurance horses. The '★' indicate significant mean difference

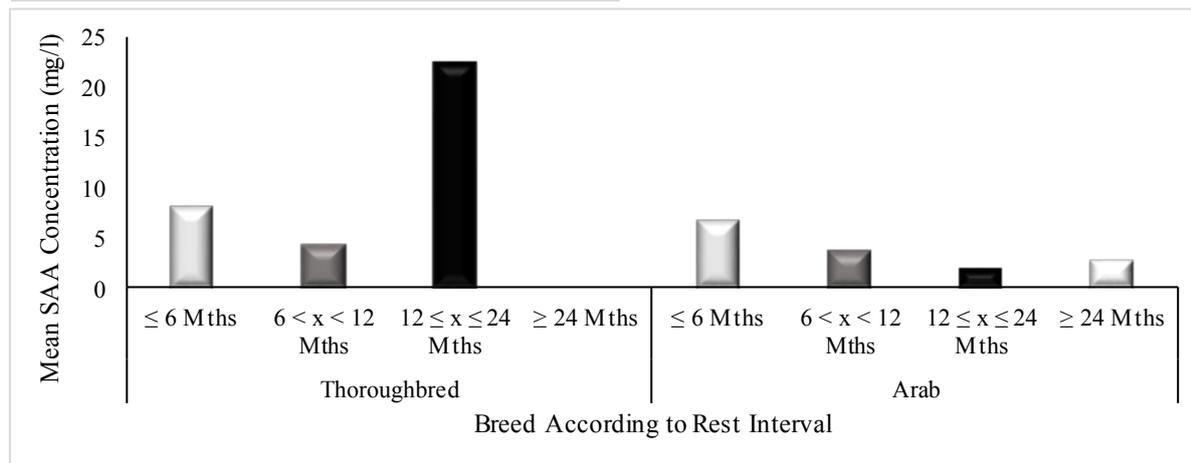


Figure 5. Comparative mean SAA concentration (mg/L) between Thoroughbred (n = 4) and Arabian (n = 36) endurance horse according to rest interval

According to age groups, the mean SAA concentrations were highest in horses aged 6 to 11 years ( $8.38 \pm 2.25$  mg/L) followed in order by horses aged >19 years ( $3.80 \pm 0.55$  mg/L) and between 12 to 18 years ( $3.28 \pm 0.42$  mg/L) (Figure 2). There was no difference in SAA concentrations between horses aged 6 to 11 years and 12 and 18 years (Figure 2).

The SAA concentrations among horses of various age groups and during the resting interval were compared. The SAA concentrations were highest in young horses age 6 to 11 years old ( $22.53 \pm 0.00$  mg/L) that participated in endurance races 12 to 24 months followed by those that raced < 6 months prior to the study (Figure 3). The SAA concentration among horses of other age and prior race participation groups differ significantly.

Mares appeared to show highest mean SAA concentration ( $7.33 \pm 1.96$  mg/L) followed by stallions ( $3.78 \pm 0.79$  mg/L) and geldings ( $3.67 \pm 0.66$  mg/L) (Figure 4). Among the mares, the group that participated in endurance races 12 s to 24 months prior to the study had highest SAA concentration ( $22.53 \pm 0.00$  mg/L), followed by those that participated ≤ 6 months ( $8.30 \pm 2.84$  mg/L), 6 to 12 months ( $4.88 \pm 0.12$  mg/L), and finally > 24 months before the study (Figure 5). In geldings, the mean SAA concentrations ( $22.53 \pm 0.00$  mg/L) were highest in those that participated in endurance ≤ 6 months before the study and decreased with duration of rest period (Figure 5).

The SAA concentrations in Thoroughbred horses ( $9.25 \pm 3.95$  mg/L) were twice as high compared to the

Arabian horses ( $4.57 \pm 0.82$  mg/L) (Figure 6). Among the Thoroughbred horses, those horses that last participated in endurance races 12 to 24 months before the study showed the highest SAA concentrations ( $23.53 \pm 0.00$  mg/L), followed in order by horses that participated ≤ 6 months ( $8.3 \pm 2.84$  mg/L), 6 to 12 months before the study (Figure 7). In Arabian horses, the SAA concentrations were highest for the horses participated ≤ 6 months ( $6.11 \pm 6.99$  mg/L), followed in order 6 to 12 months ( $3.67 \pm 1.73$  mg/L), > 24 months, and 12 to 24 months before the study (Figure 7).

## DISCUSSION

This study was undertaken to determine the level of SAA in clinically normal endurance horses during the rest period in-between races. Several factors has been taken into considerations are the effect of resting interval, age, gender, and breed on the mean SAA concentrations.

The mean SAA concentrations for horses that participated in endurance races for 12 to 24 months were high. There was one outlier in this group of horses, showing very high mean SAA concentration of  $22.50 \pm 0.00$  mg/L, while the range of SAA concentrations for the rest of horses in this group was 0.00 to 4.00 mg/L. In general, the mean SAA concentration of these horses was similar to that of horses that participated in endurance races ≤ 6 months but higher than those that participated for 6 to 12 months and > 24 months. The results suggest that the SAA concentration in endurance horses is related

to the period of participation in races. Based on the SAA concentrations levels, the group of horses in this study that participated in endurance races for durations of < 6 months and > 24 months had developed some inflammatory reactions (Cywińska *et al.*, 2013). These findings may be incidental and could also be due to orthopaedic injuries. Orthopaedic injuries in active endurance horses are common, and could be the result of tears in forelimb suspensory ligament and superficial digital flexor tendons (Gomide *et al.*, 2006) and metabolic disorders (Allen *et al.*, 1988). At this juncture, the correlation between SAA concentration and duration of participation in races before resting is evident; but, there are other factors that contributes to SAA concentrations in these horses that include fitness, intensity and method of training, race conditions and environment (Jacobsen *et al.*, 2006; 2007).

Young horses aged 6 to 11 years old horses tended to have higher mean SAA concentration than their older counterparts. Young horses are to yet to adapt to the rigors of training and competition. Thus, it is presumed that they are more prone to inflammatory reactions and injuries during the on-going process of adaptation. Older horses aged 12 to 18 years in this group are adapted to the training and fit for endurance races, thus, they showed less tendency to show inflammatory responses as indicated by the low SAA concentrations (Giori *et al.*, 2011). However, musculature loses elasticity with age, which could contribute to muscle injuries and damage during intense training and rigorous races. However, even in fit horses, the SAA concentrations also vary with age; that is middle aged horses with greater muscle elasticity are expected to show lower SAA than old horses, as suggest by the current study (Kenyon *et al.*, 2007).

The SAA concentrations in endurance horses vary with gender. For example, in this study of resting horses, geldings and stallions showed much lower mean SAA concentrations than mares. Mares are more affected by hormonal cycles than either geldings or stallions. It is expected that the oestrus cycle in mares could have contributed to the development of inflammatory responses in these horses. It was also previous shown that during proestrus and metestrus, mares tend to exhibit very aggressive and stressful behaviour, which may result in inflammatory response and subsequent increase in SAA concentration (Hultén *et al.*, 1997).

There is variability in mean SAA concentration among horse breeds. Thoroughbred horses had high SAA concentration than Arabian horses, due to composition of their fast twitch muscle fibres (Thiruvankadan *et al.*, 2009). Fast twitch muscle fibres are highly anaerobic, although provides the Thoroughbred with speed but also accumulate a lot more muscle-damaging lactate than the Arabians. Since lactate accumulation in muscles lead to exhaustion, these horses are more prone to injuries and inflammation, thus, increase in SAA concentration. Arabian horses on the other hand have slow twitch aerobic muscle fibres. These muscles generate high energy. These muscles are “fatigue-resistant” and capable of reducing the toxic end-products of metabolism. Since endurance rides are long duration submaximal intensity aerobic events, horses with slower twitch fibers are ideal for these races (Revold *et al.*, 2010). Arabian horses with these

attributes have low tendency to be affected by exertions during endurance rides. This is clearly reflected by the low SAA concentrations in the Arabian horses than the Thoroughbreds as indicated by their low SAA concentrations (Adamu *et al.*, 2014).

One of the objectives of the this study was to develop the reference range for SAA in endurance horses, The study proposed the reference range for endurance race horses to be from 2.09 to 8.01 mg/L. In fact, 77.5% of the horses in this study showed SAA concentration within this range.

## CONCLUSION

It is suggested that the reference range for SAA concentration of endurance race horses in Malaysia is from 2.09 to 8.01mg/L. It is proposed that SAA concentrations can be used as an indicator of fitness and ability of horses to complete endurance races. The owners should allow for longer rest periods for horses that show SAA concentrations exceeding the reference range.

## ACKNOWLEDGEMENTS

The gratitude goes to Dr. Shri Kanth Kaenasalingam from the Selangor Turf Club, Dr. John Tito Sapalo from Skuadron Istiadat Berkuda and Dr. Hamidah Helman from the Department of Veterinary Services Malaysia for assisting us for blood sampling of horses for this project. A special thanks to Prof. Rasedee Abdullah from the Faculty of Veterinary Medicine UPM, for the generous review and editorial assistance on the manuscript.

## CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

## FUNDINGS

This study was supported by research Universiti Putra Malaysia (Inisiatif Putra Muda) research grant (2013-2015).

## REFERENCES

- Adamu, L., Adzahan, N., Rasedee, A., and Bashir, A. (2014). Physical Parameters and Risk Factors Associated with the Elimination of Arabian and Crossed Arabian Endurance Horses during a 120-km Endurance Race. *Journal of Equine Veterinary Science*. 34(4): 494–499.
- Aldred, A.R., and Schreiber, G. (1993). The negative acute phase proteins. In: *Acute Phase Proteins. Molecular Biology, Biochemistry, and Clinical Applications*, CRC Press, Boca Raton, 21-37.
- Allen, B.V., and Kold, S.E. (1988). Fibrinogen response to surgical tissue trauma in the horse. *Journal of Veterinary Equine*. 20: 441-443.
- Cywińska, A., Szarska, E., Witkowski, L., Schollenberger, A., and Winnicka, A. (2013). Serum amyloid A ( SAA ) concentration after training sessions in Arabian race and endurance horses Serum amyloid A ( SAA ) concentration after training sessions in Arabian race and endurance horses. *BMC Veterinary Research*. 9(91): 1-7.
- Cywińska, A., Szarska, E., Goreccka, R., Witkowski, L., Hecold, M., Berenowski, A., Schollenberger, A., and Winnicka, A. (2012). Acute phase protein concentrations after limited distance and long

- distance endurance rides in horses. *Research in veterinary science*. 93(3): 1402–6.
- Gabay, C., and Kushner, I. (1999). Acute-phase proteins and other systemic response to inflammation. *Journal of New England Medicine*. 340: 448-454.
- Giori, L., Moretti, P., Giordano, A., and Paltrinieri, S. (2011). Short-term evaluation of serum amyloid a after exercise in clinically healthy horses. *Journal of Equine Veterinary Science*. 31(9): 499–501.
- Gomide, L.M., Silva, M.A., Martins, C.B., Albernaz, R.M., Orozco, C., and Lacerda- Neto, J.C. (2006). Musculoskeletal injuries of Arabian horses during endurance training. *Proceedings of Conference on Equine Sports Medicine and Science*, Cambridge, UK, 185–18.
- Hultén, C., Sletten, K., Bruun, C. F., and Marhaug, G. (1997). The acute phase serum amyloid A protein (SAA) in the horse: Isolation and characterization of three isoforms. *Veterinary Immunology and Immunopathology*. 57(3-4): 215–227.
- Jacobsen, S., and Andersen, P.H. (2007). The acute phase protein serum amyloid A (SAA) as a marker of inflammation in horses. *Equine Veterinary Education*. 19: 38–46.
- Jacobsen, S., Thomsen, M.H., and Nanni, S. (2006). Concentrations of serum amyloid A in serum and synovial fluid from healthy horses and horses with joint disease. *American Journal of Veterinary Research*. 67(10): 1738-1742.
- Kent, J., 1992. Guest editorial acute phase proteins: Their use in veterinary. *British Veterinary Journal*. 9–12.
- Kenyon, C.L., Basaraba, R.J., and Bohn, A.A. (2007). Influence of endurance exercise on serum concentrations of iron and acute phase proteins in racing sled dogs. *Journal of the American Veterinary Medicine Association*. 239(9): 1201–1210.
- Koj, A. (1996). Initiation of acute phase response and synthesis of cytokines. *Biochimica et Biophysica Acta*. 1317: 84-94.
- Kushner, I., and Mackiewicz, A. (1993). The acute phase response: an overview. In: *Acute Phase Proteins. Molecular Biology, Biochemistry, and Clinical Applications*, CRC Press, Boca Raton, Florida, 3-19.
- Mozes, G., Friedman, N., and Shainkin-Kestenbaum, R.(1989). Serum amyloid A: An extremely sensitive marker for intensity of tissue damage in trauma patients and indicator of acute phase response in various diseases. *Journal of Trauma*. 29: 71-74.
- Nagy, A., Dyson, S.J., and Murray, J.K. (2012). A veterinary review of endurance riding as an international competitive sport. *Veterinary Journal*. 194(3): 288–93.
- Pepys, M.B., and Baltz, M.L. (1983). Acute phase proteins with special reference to C-Reactive Protein and related proteins (Pentaxins) and Serum Amyloid A protein. *Advances in Immunology*, 34, 141-212 .
- Revolv, T., Mykkänen, A., Karlström, K., Ihler, C., Pösö, A. and Essén-Gustavsson, B. (2010). Effects of training on equine muscle fibres and monocarboxylate transporters in young Coldblooded Trotters. *Equine Veterinary Journal*. 42: 289-295.
- Thiruvankadan, A.K., Kandasamy, N., and Panneerselvam, S. (2009). Inheritance of racing performance of Thoroughbred horses. *Livestock Science*. 121(2-3): 308–326.
- Uhlar, C.M., and Whitehead, A.S. (1999). Serum amyloid A, the major vertebrate acute-phase reactant. *European Journal of Biochemistry*. 265: 501-523.